## Non-unique Games over Compact Groups and Orientation Estimation in Cryo-EM

#### Amit Singer

Princeton University

#### Department of Mathematics and Program in Applied and Computational Mathematics

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National Institute of General Medical Sciences

Basic Discoveries for Better Health







Amit Singer (Princeton University)

## Single Particle Reconstruction using cryo-EM

Schematic drawing of the imaging process:



The cryo-EM problem:



### New detector technology: Exciting times for cryo-EM

www.sciencemag.org SCIENCE VOL 343 28 MARCH 2014

#### BIOCHEMISTRY

### **The Resolution Revolution**

Werner Kühlbrandt

recise knowledge of the structure of macromolecules in the cell is essential for understanding how they function. Structures of large macromolecules can now be obtained at near-atomic resolution by averaging thousands of electron microscope images recorded before radiation damage accumulates This is what Amunts et al have done in their research article on page 1485 of this issue (1), reporting the structure of the large subunit of the mitochondrial ribosome at 3.2 Å resolution by electron crvo-microscopy (cryo-EM). Together with other recent high-resolution crvo-EM structures (2-4) (see the figure), this achievement heralds the beginning of a new era in molecular biology. where structures at near-atomic resolution are no longer the prerogative of x-ray crystallography or nuclear magnetic resonance (NMR) spectroscopy.

Ribosomes are ancient, massive protein-RNA complexes that translate the linear genetic code into three-dimensional proteins. Advances in detector technology and image processing are yielding high-resolution electron cryo-microscopy structures of biomolecules.



Near-atomic resolution with cryo-EM. (A) The large subunit of the yeast mitochondrial ribosome at 3.2 Å reported by Amunts et al. In the detailed view below, the base pairs of an RNA double helix and a magnesium ion (blue) are clearly resolved. (B) TRV1 ion channel at 3.4 Å (2), with a detailed view of residues liming the ion pore on the four-fold axis of the tetrameric channel. (O  $\Gamma_{nor}$ -reducing [NiFe] hydrogenase at 3.3 Å  $\delta$  (3). The detail shows an ch kix in the FrlA subunit with resolved side chains. The magna en ot drawn to scale.

#### Cryo-EM in the news...

March 31, 2016

Science

REPORTS

Cite as: Sirohi et al., Science 10.1126/science.aaf5316 (2016).

#### The 3.8 Å resolution cryo-EM structure of Zika virus

#### Devika Sirohi,<sup>1\*</sup> Zhenguo Chen,<sup>1\*</sup> Lei Sun,<sup>1</sup> Thomas Klose,<sup>1</sup> Theodore C. Pierson,<sup>2</sup> Michael G. Rossmann,<sup>1</sup>† Richard J. Kuhn<sup>1</sup>†

<sup>1</sup>Markey Center for Structural Biology and Purdue Institute for Inflammation, Immunology and Infectious Disease, Purdue University, West Lafayette, IN 47907, USA. <sup>2</sup>Viral Pathogenesis Section, Laboratory of Viral Diseases, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda, MD 20892, USA.

#### Method of the Year 2015

January 2016 Volume 13 No 1



Single-particle cryo-electron microscopy (cryo-EM) is our choice for Method of the Year 2015 for its newfound ability to solve protein structures at near-atomic resolution. Featured is the 2.2-Å cryo-EM structure of β-galactosidase as recently reported by Bartesaghi et al. (Science 348, 1147–1151, 2015). Cover design by Erin Dewalt.

#### Big "Movie" Data, Publicly Available

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Dataset 🌢	Title	Authors \$	Related EMDB/PDB \$ entries	Size 🗸	Marabin R et al. (2016) The Gurgeas Bioinformatics Institute in 2016 D ata growth and integration. Code C E et al. (2016) CrywB1 single particle analysis with the volta phase plane. Darev R, Baumelster W. (2016) Particle alignment reliability in single particle level con overvice/codexy a general
EMPLAR-10051	2.2 A resolution cryo-EM structure of beta-galactosidase in complex with a cell-permeant inhibitor [multiple data sets in MRC format]	Bartesaghi A. Mark A. Banerjee S. Matthies D. Wu X. Mins JL Subramaniam S [Pubmed: <u>29593817]</u> [D02: <u>10.1126/science.asb1576]</u>	EMD-2984, Sala	12.4 78	
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EMPIAR-10023	Electron cryo-microscopy of ATP synthase dimers from Polycomella sp. [2823 multi-frame micrographs composed of 24 frames each in MRC format]	Allegretti M, Klusch N, Mills DJ, Vonck J, Kuehlorandt W, Davkes KM [Pubmed: 25707805] [DOI: 10.1038/insture14183]	EMD-2852	4.1 TB	

## E. coli 50S ribosomal subunit

27,000 particle images provided by Dr. Fred Sigworth, Yale Medical School



#### 3D reconstruction by S, Lanhui Wang, and Jane Zhao

## Orientation Estimation: Fourier projection-slice theorem



- Cryo-EM inverse problem: Find  $\phi$  (and  $R_1, \ldots, R_n$ ) given  $I_1, \ldots, I_n$ .
- n = 3: Vainshtein and Goncharov 1986, van Heel 1987
- n > 3: S, Shkolnisky (SIAM Imaging 2011)

$$\min_{R_1, R_2, \dots, R_n \in SO(3)} \sum_{i \neq j} \|R_i c_{ij} - R_j c_{ji}\|^2$$

### Maximum Likelihood Estimation

- The images contain more information than that expressed by optimal pairwise matching of common lines.
- Algorithms based on pairwise matching can succeed only at "high" SNR.
- We would like to try all possible rotations  $R_1, \ldots, R_n$  and choose the combination for which the agreement on the common lines (implied by the rotations) as observed in the images is maximal.
- Computationally intractable: exponentially large search space, complicated cost function.

$$\min_{g_1,\ldots,g_n\in G}\sum_{i,j=1}^n f_{ij}(g_ig_j^{-1})$$





G = SO(3)

$$\min_{g_1,g_2,...,g_n\in G} \sum_{i,j=1}^n f_{ij}(g_i g_j^{-1})$$

#### Non-Unique Games over Compact Groups

• Optimization problem:

$$\min_{g_1,g_2,...,g_n\in G} \sum_{i,j=1}^n f_{ij}(g_ig_j^{-1})$$

*G* is a compact group,  $f_{ij}: G \to \mathbb{R}$  smooth, bandlimited functions.

- Parameter space  $G \times G \times \cdots \times G$  is exponentially large.
- For  $G = \mathbb{Z}_2 = \{-1, +1\}$  this encodes Max-Cut, which is NP-hard.



#### Why non-unique games?

 Max-2-Lin(ℤ<sub>L</sub>) formulation of Unique Games (Khot et al 2005): Find x<sub>1</sub>,..., x<sub>n</sub> ∈ ℤ<sub>L</sub> that satisfy as many difference eqs as possible

$$x_i - x_j = b_{ij} \mod L, \quad (i,j) \in E$$

• This corresponds to  $G = \mathbb{Z}_L$  and

$$f_{ij}(x) = \left\{ egin{array}{cc} -1 & x = b_{ij} \ 0 & x 
eq b_{ij} \end{array} 
ight.$$

in

$$\min_{x_1,x_2,\ldots,x_n\in\mathbb{Z}_L}\sum_{i,j=1}^n f_{ij}(x_i-x_j)$$

 Our games are non-unique in general, and the group is not necessarily finite.

#### Fourier transform over G

• Recall for G = SO(2)

$$f(\alpha) = \sum_{k=-\infty}^{\infty} \hat{f}(k) e^{ik\alpha}$$
$$\hat{f}(k) = \frac{1}{2\pi} \int_{0}^{2\pi} f(\alpha) e^{-ik\alpha} d\alpha$$

• In general, for a compact group G

$$f(g) = \sum_{k=0}^{\infty} d_k \operatorname{Tr} \left[ \hat{f}(k) \rho_k(g) \right]$$
$$\hat{f}(k) = \int_G f(g) \rho_k(g)^* dg$$

- Here
  - $\rho_k$  are the unitary irreducible representations of G
  - d<sub>k</sub> is the dimension of the representation ρ<sub>k</sub> (e.g., d<sub>k</sub> = 1 for SO(2), d<sub>k</sub> = 2k + 1 for SO(3))
  - dg is the Haar measure on G

#### Linearization of the cost function

Introduce matrix variables ("matrix lifting")

$$X_{ij}^{(k)} = \rho_k(g_i g_j^{-1})$$

• Fourier expansion of f<sub>ij</sub>

$$f_{ij}(g) = \sum_{k=0}^{\infty} d_k \operatorname{Tr}\left[\hat{f}_{ij}(k)\rho_k(g)\right]$$

Linear cost function

$$f(g_1, \dots, g_n) = \sum_{i,j=1}^n f_{ij}(g_i g_j^{-1}) = \sum_{i,j=1}^n \sum_{k=0}^\infty d_k \operatorname{Tr} \left[ \hat{f}_{ij}(k) X_{ij}^{(k)} \right]$$

# Constraints on the variables $X_{ii}^{(k)} = \rho_k(g_i g_i^{-1})$

$$X^{(k)} \succeq 0$$

$$X^{(k)}_{ii} = I_{d_k}, \text{ for } i = 1, ..., n$$

$$\text{rank}(X^{(k)}) = d_k$$

$$X^{(k)}_{ij} = \rho_k(g_i g_j^{-1}) = \rho_k(g_i)\rho_k(g_j^{-1}) = \rho_k(g_i)\rho_k(g_j)^*$$

$$X^{(k)} = \begin{bmatrix} \rho_k(g_1) \\ \rho_k(g_2) \\ \vdots \\ \rho_k(g_n) \end{bmatrix} \begin{bmatrix} \rho_k(g_1)^* & \rho_k(g_2)^* & \cdots & \rho_k(g_n)^* \end{bmatrix}$$

- We drop the non-convex rank constraint.
- The relaxation is too loose, as we can have  $X_{ij}^{(k)} = 0$  (for  $i \neq j$ ).
- Even with the rank constraint, nothing ensures that  $X_{ij}^{(k)}$  and  $X_{ij}^{(k')}$  correspond to the same group element  $g_i g_i^{-1}$ .

# Additional constraints on $X_{ii}^{(k)} = \rho_k(g_i g_i^{-1})$

• The delta function for G = SO(2)

$$\delta(lpha) = \sum_{k=-\infty}^{\infty} e^{iklpha}$$

• Shifting the delta function to  $\alpha_i - \alpha_j$ 

$$\delta(\alpha - (\alpha_i - \alpha_j)) = \sum_{k=-\infty}^{\infty} e^{ik\alpha} e^{-ik(\alpha_i - \alpha_j)} = \sum_{k=-\infty}^{\infty} e^{ik\alpha} X_{ij}^{(k)*}$$

• The delta function is non-negative and integrates to 1:

$$\sum_{k=-\infty}^{\infty} e^{ik\alpha} X_{ij}^{(k)*} \ge 0, \quad \forall \alpha \in [0, 2\pi)$$
$$\frac{1}{2\pi} \int_{0}^{2\pi} \sum_{k=-\infty}^{\infty} e^{ik\alpha} X_{ij}^{(k)*} d\alpha = X_{ij}^{(0)*} = 1$$

#### Finite truncation via Fejér kernel

- In practice, we cannot use infinite number of representations to compose the delta function.
- Simple truncation leads to the Dirichlet kernel which changes sign

$$D_m(\alpha) = \sum_{k=-m}^m e^{\imath k \alpha}$$

- This is also the source for the Gibbs phenomenon and the non-uniform convergence of the Fourier series.
- The Fejér kernel is non-negative

$$F_m(\alpha) = \frac{1}{m} \sum_{k=0}^{m-1} D_k(\alpha) = \sum_{k=-m}^m \left(1 - \frac{|k|}{m}\right) e^{ik\alpha}$$

• The Fejér kernel is the first order Cesàro mean of the Dirichlet kernel.

#### Finite truncation via Fejér-Riesz factorization

• Non-negativity constraints over *SO*(2)

$$\sum_{k=-m}^{m} \left(1 - \frac{|k|}{m}\right) e^{\imath k \alpha} X_{ij}^{(k)^*} \ge 0, \quad \forall \alpha \in [0, 2\pi)$$

- Fejér-Riesz: P is a non-negative trigonometric polynomial over the circle, i.e. P(e<sup>iα</sup>) ≥ 0 ∀α ∈ [0, 2π) iff P(e<sup>iα</sup>) = |Q(e<sup>iα</sup>)|<sup>2</sup> for some polynomial Q.
- Leads to semidefinite constraints on  $\left\{X_{ij}^{(k)}\right\}_{i}$  for each i, j.
- Similar non-negativity constraints hold for general G using the delta function over G

$$\delta(g) = \sum_{k=0}^{\infty} d_k \operatorname{Tr} \left[ \rho_k(g) \right]$$

• For example, Fejér proved that for *SO*(3) the second order Cesàro mean of the Dirichlet kernel is non-negative.

• 
$$X_{ij}^{(k)}$$
 is a representation of  $SO(3)$ .

• 
$$X_{ij}^{(0)} = 1$$

 X<sup>(1)</sup><sub>ij</sub> ∈ convSO(3) is a semidefinite constraint using unit quaternions and Euler-Rodrigues formula (Saunderson, Parrilo, Willsky SIOPT 2015)

$$Q = qq^T$$
:  $Q \succeq 0$ ,  $\operatorname{Tr}(Q) = 1$ ,  $Q = T(X_{ij}^{(1)})$ 

•  $X_{ij}^{(k)}$  sum-of-squares relaxation

#### Tightness of the semidefinite program

- We solve an SDP for the matrices  $X^{(1)}, \ldots, X^{(m)}$ .
- Numerically, the solution of the SDP has the desired ranks up to a certain level of noise (w.h.p).
- In other words, even though the search-space is exponentially large, we typically find the MLE in polynomial time.
- This is a viable alternative to heuristic methods such as EM and alternating minimization.
- The SDP gives a certificate whenever it finds the MLE.

#### **Final Remarks**

• Loss of handedness ambiguity in cryo-EM: If  $g_1, \ldots, g_n \in SO(3)$  is the solution, then so is  $Jg_1J^{-1}, \ldots, Jg_nJ^{-1}$  for J = diag(-1, -1, 1).

• Define 
$$X_{ij}^{(k)} = \frac{1}{2} \left[ \rho_k(g_i g_j^{-1}) + \rho_k(Jg_i g_j^{-1} J^{-1}) \right]$$

- Splits the representation: 2k + 1 = d<sub>k</sub> = k + (k + 1), reduced computation
- Point group symmetry (cyclic, dihedral, etc.): reduces the dimension of the representation (invariant polynomials)
- Translations and rotations simultaneously: *SE*(3) is a non-compact group, but we can map it to *SO*(4).
- Simultaneous rotation estimation and classification

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## ASPIRE: Algorithms for Single Particle Reconstruction

Open source toolbox, publicly available:

http://spr.math.princeton.edu/



#### **About Our Project**

Our project aims to develop mathematical theory and improved algorithms for 3D molecular structure determination using cryo-EM. Our methods use a combination of tools from tomography, convex optimization and semidefinite programming, random matrix theory, signal and image processing, statistics, machine learning, nonlinear dimensionality reduction, randomized algorithms in numerical linear lagebra, and representation theory.

Acknowledgement: This project would not have been possible without the support of Award Number R01GM090200 from the National Institute of General Medical Sciences (NIGMS). The content is solely the responsibility of the authors and does not necessarily represent the official views of the NIGMS or the NIH.

What is cryo-EM and SPR? "Three dimensional electron microscopy" is the name commonly given to methods in which the 3D structures of macromolecular complexes are obtained from sets of 2D projection images taken in an electron microscope. The most widespread and general of these methods is single-particle reconstruction (SPR). In SPR, It also structure is determined from images of randomly oriented and positioned, identical macromolecular 'arriteles', typically complexes S00 kDa or larger in size. The SPR method has been applied to images of nadomly oriented and positioned, identical macromolecular 'stricles', typically complexes S00 kDa or larger in size. The SPR method has been applied to images of nagatively stained specimens, and to images obtained from frozen-hydrated, unstained specimens. In the latter technique, called cryo-EM, the sample of macromolecules is rapidly frozen in thin (-100 nm) layer of vitreous ice, and maintained at liquid nitrogen temperature throughout the imaging process. SR from cryo-SR from cryo-EM images is of particular interest because it promises to be an entirely general technique. I does not require